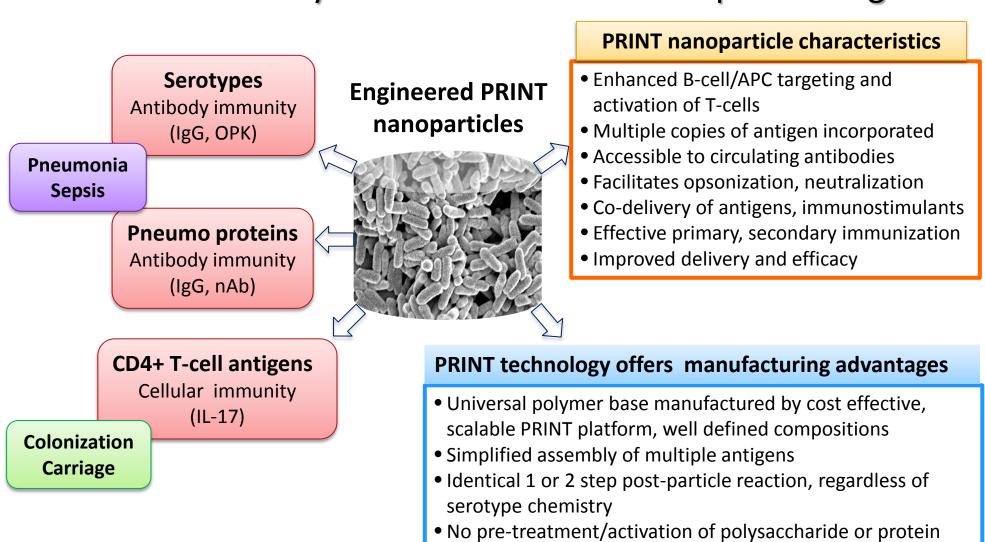
Next Generation Multivalent PRINT® Nanoparticle Vaccine Targeting Pneumococcal Disease

Presenter: Gabe Fawcett (gabe.fawcett@liquidia.com)

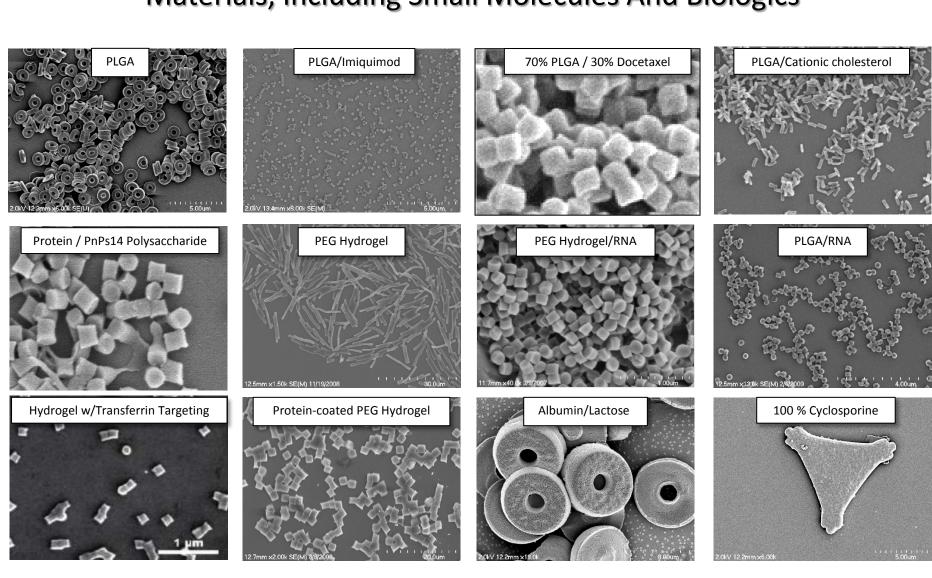
Authors: Shyam Rele, RiLee Robeson, Anton Beletskii, Jeremy Hansen, Meredith Earl, Gabe Fawcett, Marquita Lilly, Joseph Marchand, Michele Stone, Camille Bernasconi, Jinny Conley, Michael Hunter, Ramya Yadavalli, Nicole Meyer, Lara Kelly, Ben Yerxa,¹ Jeff Maisonneuve,² Mark Alderson,² Frank Malinoski¹

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PRINT Technology Offers The Flexibility To Target Humoral And Cellular Immunity Towards Pneumococcal Specific Targets



PRINT Is Compatible With Numerous Types Of Pharmaceutical Materials, Including Small Molecules And Biologics



In vitro ELISA and DIONEX HPLC methods to quantify

polysaccharide levels in PRINT nanoparticle formulations

≥ 240 220

180

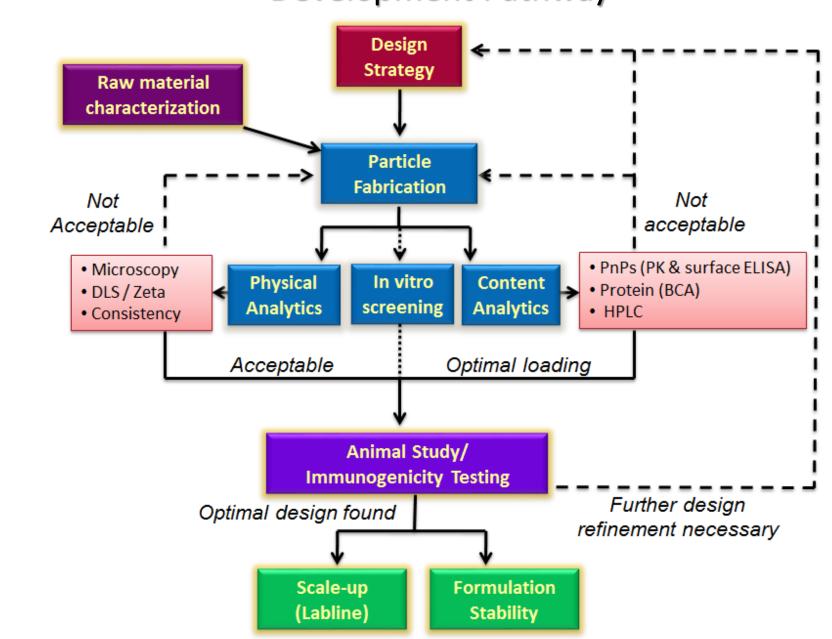
PnPs5

2.00

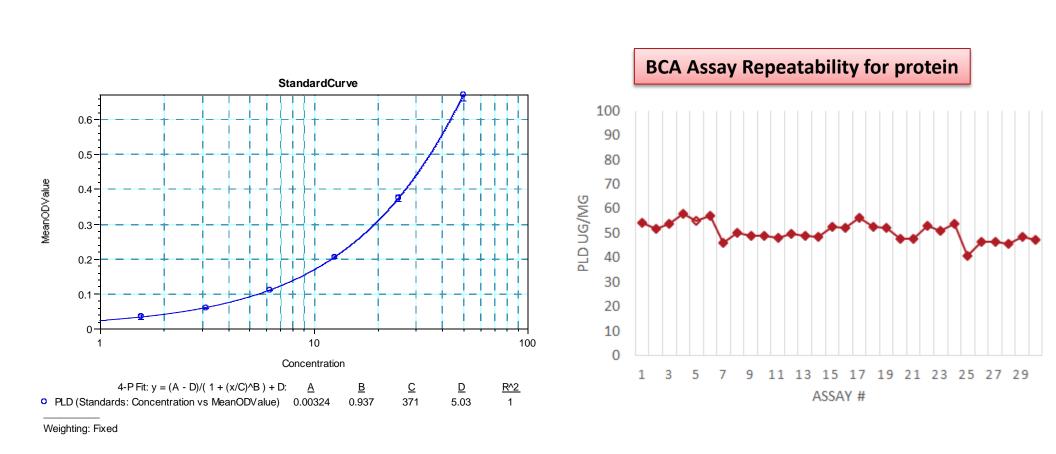
4.00

PnPs6A

PRINT Pneumococcal Polysaccharide Vaccine **Development Pathway**



Protein in PRINT formulations can be reliably quantified in solid phase using a BCA colorimetric assay



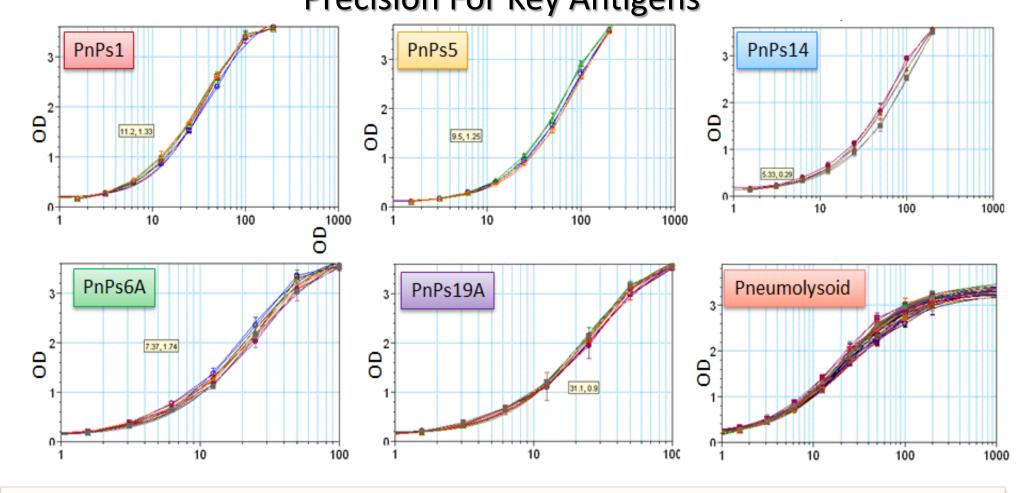
- Versatile method allows quantification of multiple antigenic carrier proteins in solid phase
- Good assay-to-assay precision for protein analysis in nanoparticles

PLY Neutralization Reciprocal Titer

Correlation between

• No pre-treatment/activation of polysaccharide or protein Ease of purification from unreacted protein, polysaccharide Sterilization by filtration

In-house IgG ELISA Shows Excellent Reproducibility And **Precision For Key Antigens**



- The IgG ELISAs were adapted using standard W.H.O. IgG ELISA protocol
- PnPs specific IgG ELISAs have been adapted to all Prevnar13 serotypes
- Pneumococcal protein specific IgG ELISA also developed

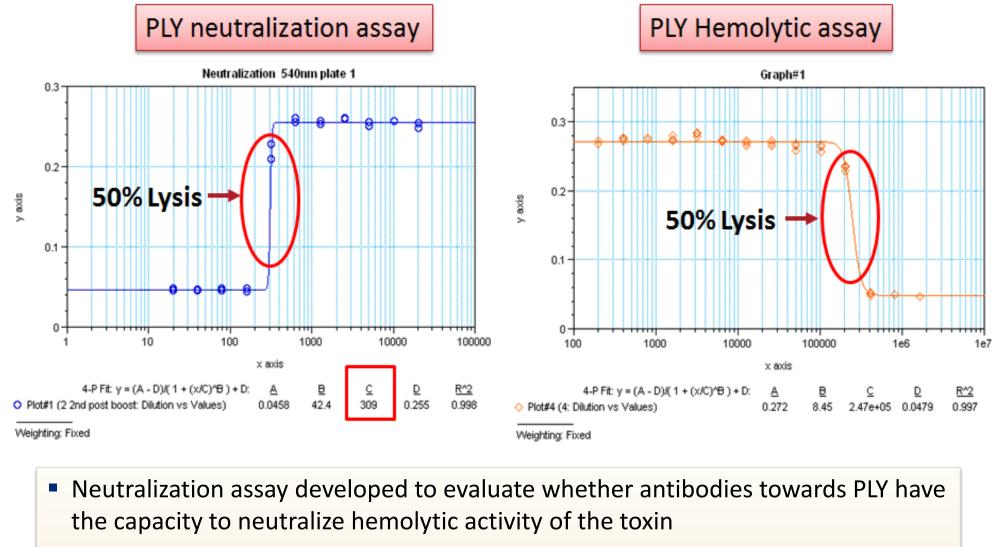
Snapshot of Assays Developed for Multi-Serotype PRINT Pneumococcal Vaccine

Serotype/ Antigen	PnPs, Protein Detection in PRINT (ELISA, DIONEX, BCA)	IgG ELISA	<i>In Vivo</i> Evaluation (IgG, OPK, nAb)	
1	€	€	mice, rabbits	W
3	⊘	⊘		
4	⊘	⊘	mice	
5	⊘	€	mice, rabbits	
6A	⊘	€	mice	W
6B	⊘	⊘		
7F	lacksquare	€		
9V				
14	⊘	\bigcirc	mice, rabbits	
18C				
19A	lacksquare	⊘	mice	
19F	€	€		
23F	⊘	€	mice	
PLD	⊘	€	mice, rabbits	
PspA	€	⊘	mice	

- •~9 Pneumococcal antigens successfully formulated with PRINT particles and evaluated in vivo to date
- Simplified and flexible assembly of antigens with PRINT particles translates easily into assay development

Functional Assessment of Pneumolysin mutant (PLD) in an in vitro toxin neutralization assay

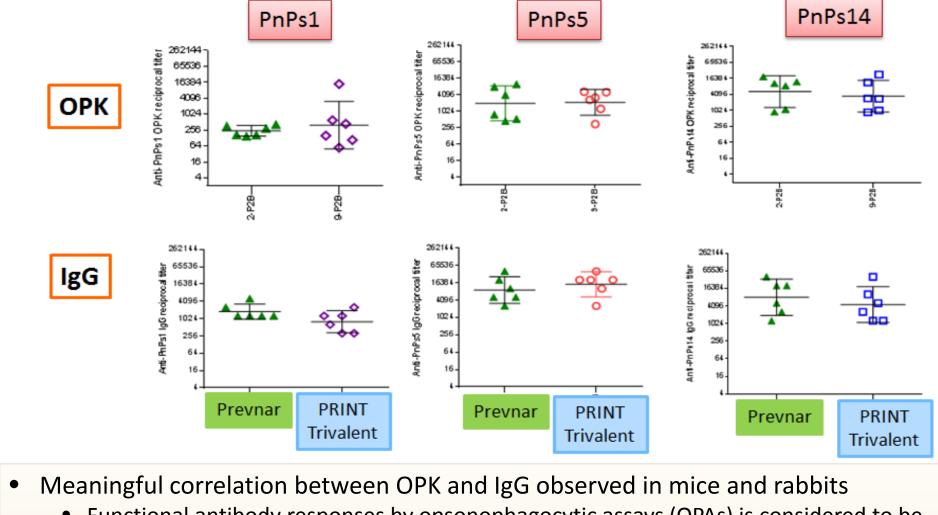
Developed ELISA and DIONEX assays for all Prevnar13 serotypes



- Hemolysis inhibition (HI) assay of PLD was developed and harmonized between multiple sites/PATH collaborators

PLY nAB and Anti-PLD IgG Robust PLD IgG antibodies were generated and shown to be functional by neutralizing hemolytic activity of PLY Strong correlation seen **PLD IgG Reciprocal Titer** between PLY neutralization titers and PLD IgG ELISA Units/mL $R^2 = 0.831$ Controls Particle Groups

Lyophilization of PRINT formulations does not diminish the

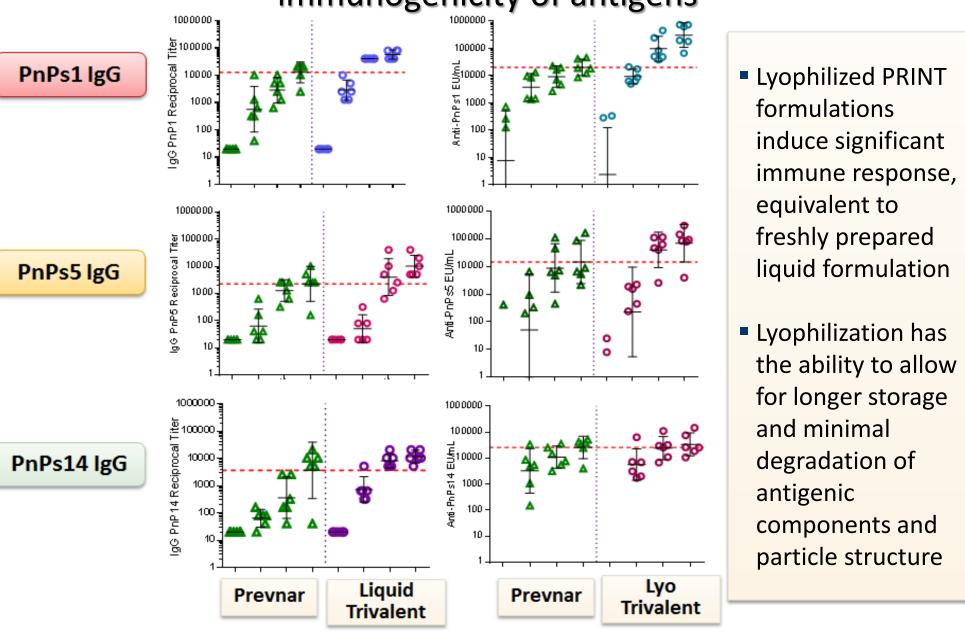


Trivalent PRINT Pneumococcal Polysaccharide Vaccine Elicits

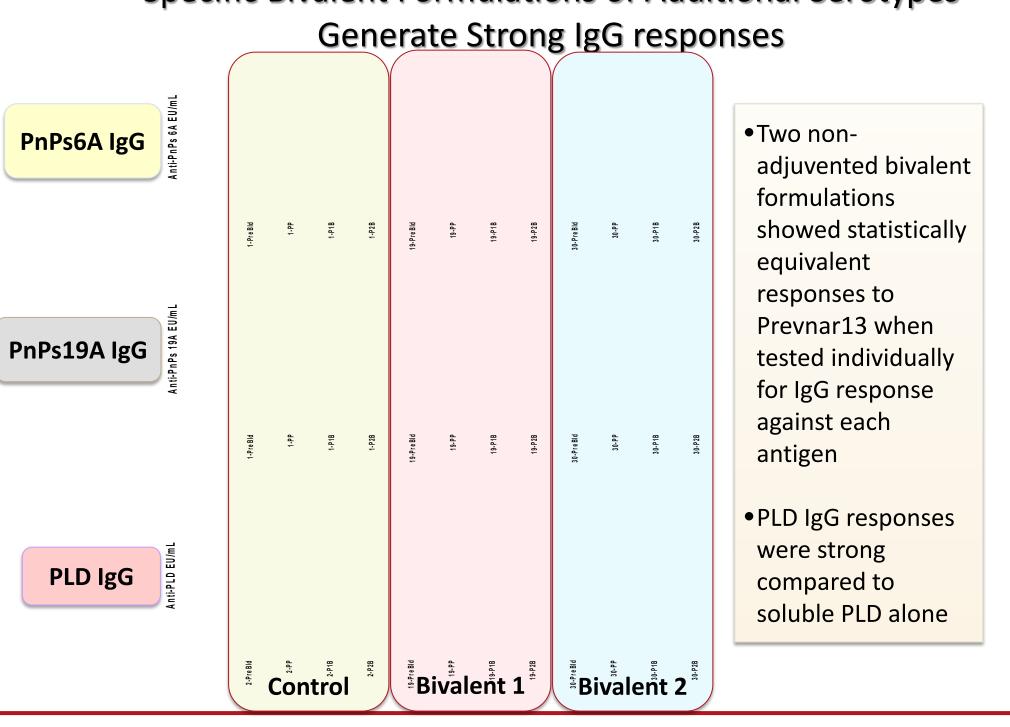
Functional Antibody Response in Rabbits

- Functional antibody responses by opsonophagocytic assays (OPAs) is considered to be useful as a surrogate marker for protection
- Sera from mice vaccinated with licensed vaccine and PRINT formulations were analyzed by OPK assay in collaboration with Dr. Moon Nahm, University of Alabama

immunogenicity of antigens



Specific Bivalent Formulations of Additional Serotypes Generate Strong IgG responses



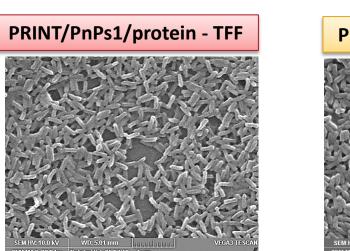
PRINT 80x80x320 nm vaccine formulations are sterile filterable (80x320 nm PRINT particles used for Influenza Clinical Program)

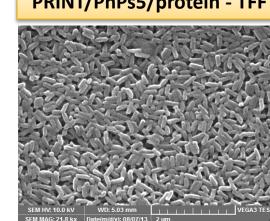
Characteristics of filtered PRINT nanoparticle dispersions (Early Trends) PnPs loading efficiency 25 -50% (optimization ongoing) (serotype, process dependent) Protein loading efficiency Generally, high protein content (serotype, process dependent) Filtration Recovery Yield > 65% DLS (nm) / PDI 300-600 nm / 0.2-0.4 Surface charge +10 to -15 mV (design, process, serotype dependent)

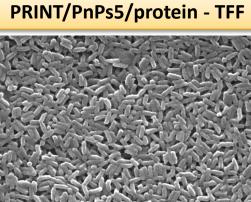
•PRINT technology enables process scale-up and allows for formulation activities to be integrated during TFF

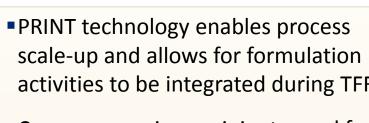
Common vaccine excipients used for filtration (polyvinyl alcohol, mannitol, sucrose, glucose, trehalose)

Process parameters and scale-up activities currently ongoing









PRINT/PnPs14/protein - TFF

PRINT Manufacturing Advantages

• Quality by Design:

- Unprecedented ability to independently control size, shape and composition
- Uniform particle population, easily characterized
- Scalable Manufacturing Platform:
 - Proprietary tooling process enables translation of patterned Silicon wafer to patterned FCR mold
 - Leveraging the breadth of Roll-to-Roll technology - adapting, not inventing machine scaling
- Commercially Relevant Scale and Cost • Minimal Facilities Burden:
 - Small footprint, low CapEx equipment (potential) for multiple, dedicated machines)

Low bioburden process, minimal water needs

No exotic utilities (drop-in-place equipment)

- Concept for aseptic manufacturing, if required
- 265 ± 10 nm
- PRINT platform demonstrates a wide-ranging multi-antigen formulation, manufacturing and analytical capability
- Robust analytical toolbox and infrastructure developed to quantify PRINT formulations antigen, protein content and IgG ELISAs (mice, rabbits) for all Prevnar13 serotypes
- Functional OPK assay test results strongly consistent with ELISA results
- Neutralizing antibody and IgG responses on par with soluble control; meaningful correlation between nAb and IgG trends
- Demonstrated the effectiveness of downstream sterile filterability and lyophilization of PRINT formulations
- The Print manufacturing process allows for rational design of next generation multivalent vaccines to address developing world needs.

