LC-SWATH/MS Metabolomics Platform with Hyphenation of Extraction for the Analysis of Polar and Non-polar Metabolites in Biological Samples

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Overview

Introduction

- Fully automated robotic sample preparation with hyphenated dual LC-SWATH/MS analysis for lipids and polar metabolites
- Automated SPE based lipid fractionation to equalize lipid abundances and reduce of runtimes
- SWATH/MS acquisition to record all the product ions originating from specific precursor ion windows in order to measrue lipids and polar metabolites
- Automated library search for polar metabolites using an in-house accurate mass MS/MS library

Evaporation Injectors device LC 30 AD Pumps Vortex Centrifuge mixer CID Q1 filter **LC Columns** Solvents Robotic Tool Change SWATH/MS (RTC) 5600 TTOF

Metabolomics studies are still challenging and technically demanding. Both sample preparation and data acquisition remain the bottlenecks in the measurements. On one hand, due to the large chemical space of the analytes and their broad dynamic range that need to be dealt with. On the other hand, metabolomics still suffer from limited selectivity due to the structural similarities within the low molecular weight compounds.

SWATH/MS opened new possibilities within Data Independent Aquisition (DIA) to gain selectivity for -omics studies as all fragments form all precursors in different isolation windows are recorded. The hyphenated setup allows full automation from sample preparation over chromatography to MS data acquisition and feature annotation. The integrated dual LC configuration with three different mobile phases enables a broad coverage of the metabolome. Additionally, the online SPE based lipid fractionation allows to equalize the dynamic range of the abundance of lipid classes.

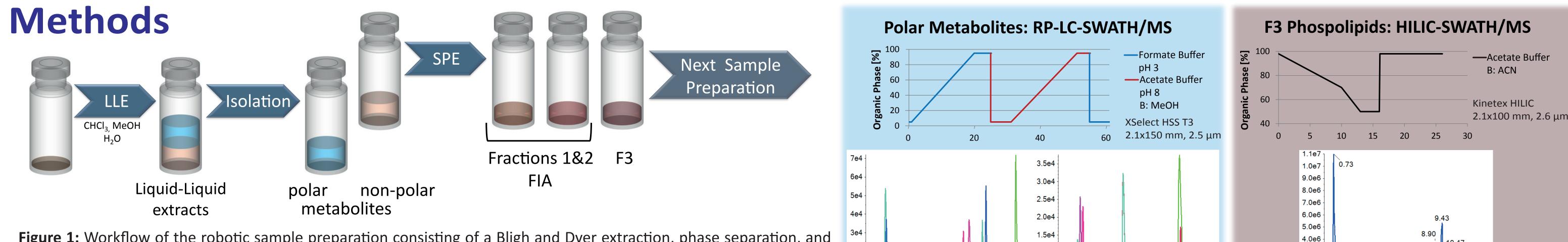
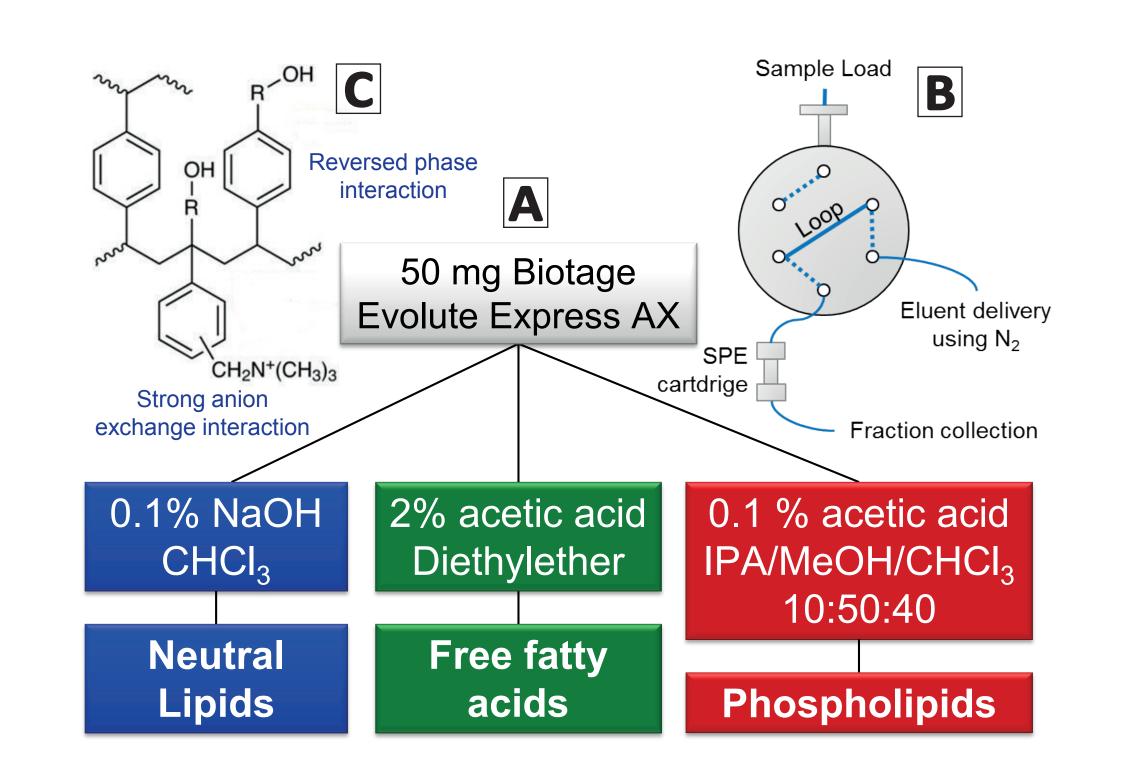
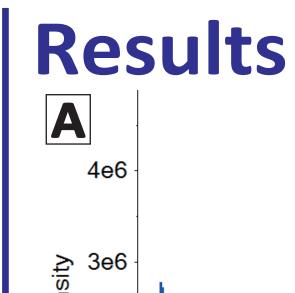


Figure 1: Workflow of the robotic sample preparation consisting of a Bligh and Dyer extraction, phase separation, and SPE using the RTC with hyphenation to a dual LC-SWATH/MS system.







3.05 3.60

Time, min

3.0e6

2.0e6

1.0e6 0.0e0

1.0e4

5.0e3

1 2 3

5678

Time, min

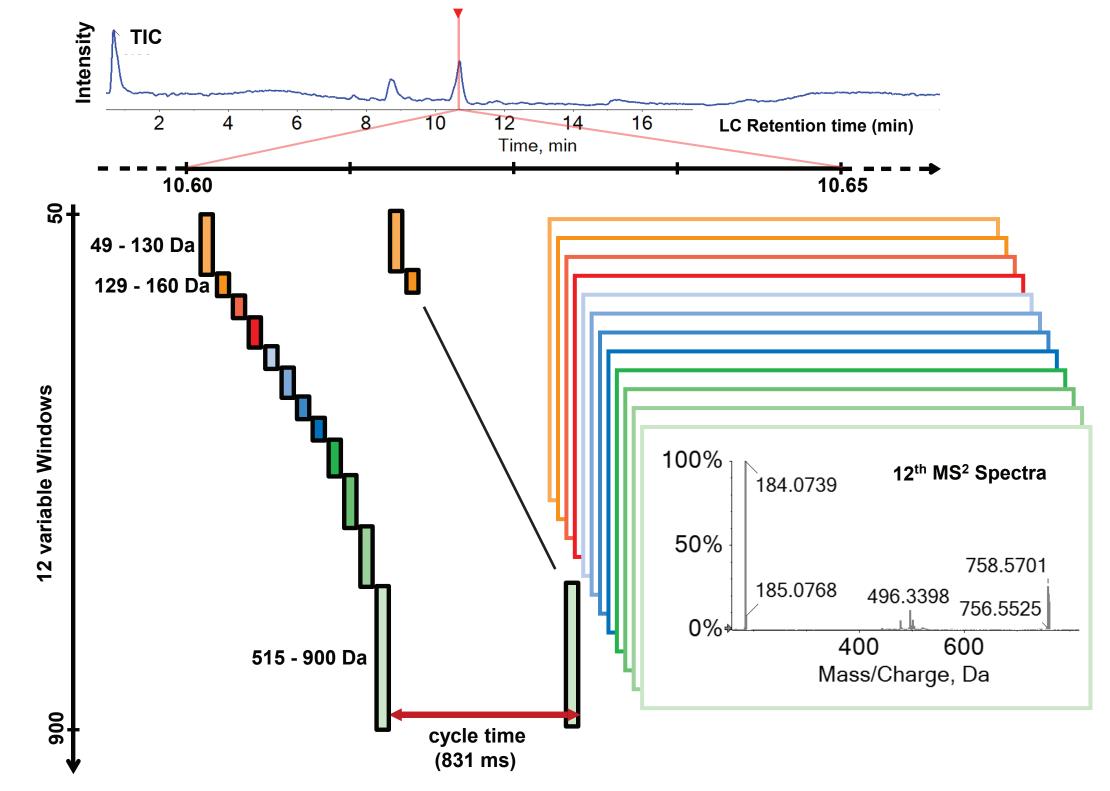
8 9 10

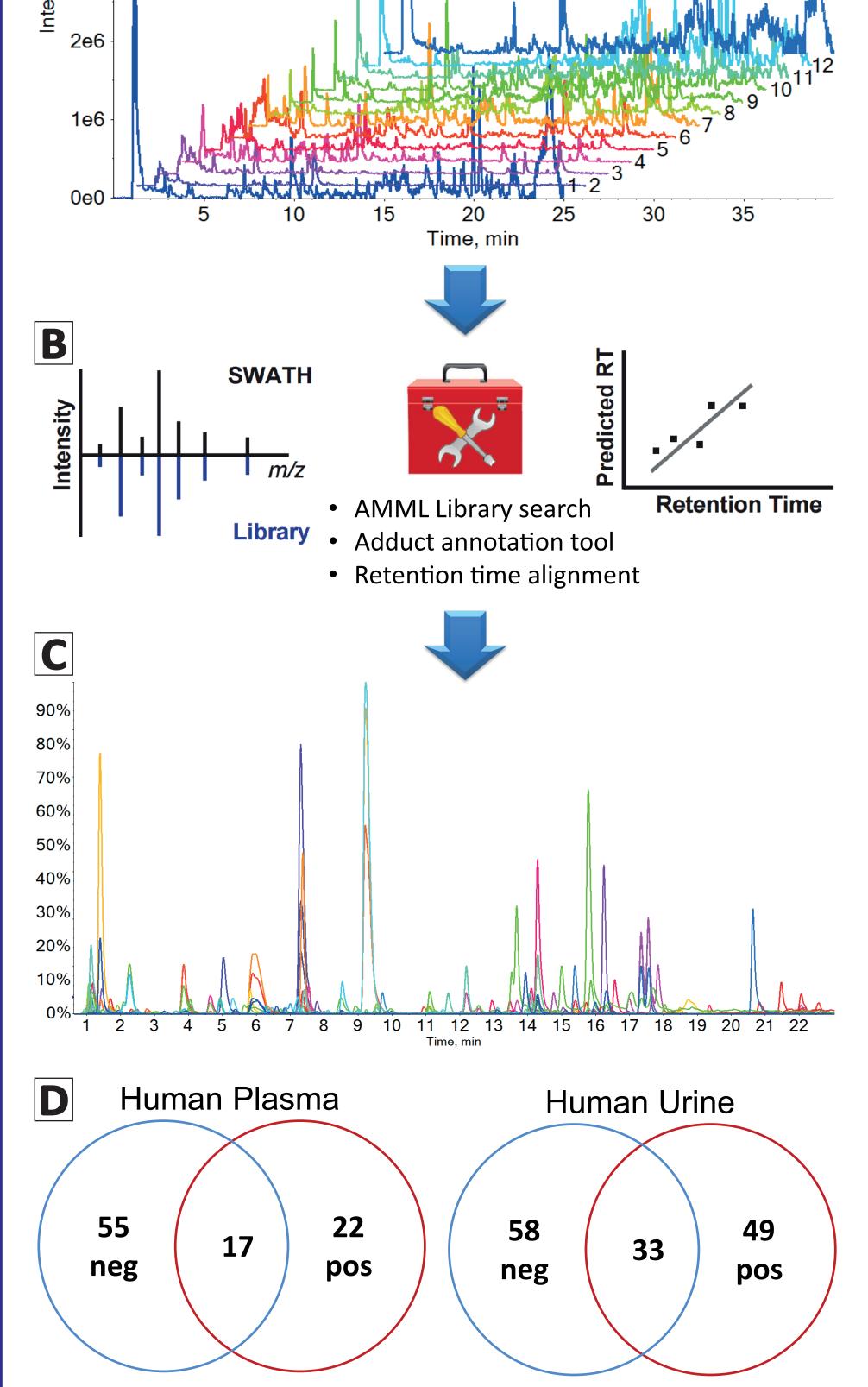
4 5 6 7

Time, min

1 2 3

Figure 2: A – Experimental setup of the automated SPE fractionation of the lipid classes. B – Injection valve system used for the elution of the fractions from the self-packed cartridge by a stream of nitrogen as carrier. C – Mixed mode sorbent used to pack the cartridge.





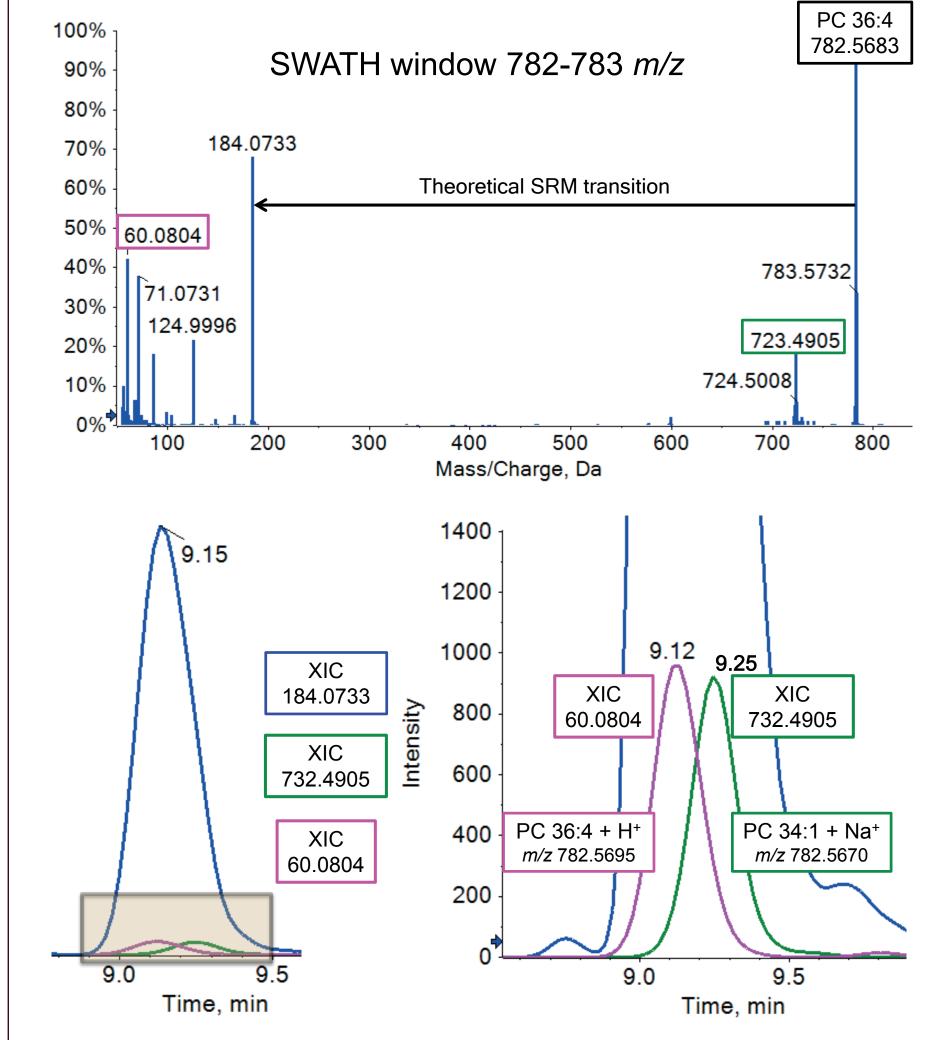


Figure 5: Top - MS2 spectrum from the SWATH Q1 window from 782-783 *m/z*. The traditionally used SRM transition from precursor to headgroup fragment ion is indicated. **Bottom –** XIC traces of selected fragment ions originating from co-eluting isobaric precursor

Figure 3: SWATH/MS setup applied for the analysis of polar metabolites in negative ion mode. Data were acquired on a Sciex 5600 TripleTOF. Parameters: 500 °C, 5.2/-4.5 kV, DP 80, 12 SWATH windows with variable unit sizes created using swathTUNER, 831 ms cycle time, 50 ms accumulation time. ions (Δm 25 mmu).

Figure 4: A – Offset chromatogram of the 12 SWATH experiments for an undiluted human urine sample (5 μ L injection volume). **B** – Inhouse tools for analyte indentification. **C** – Chromatogram of the identified analytes fulfilling the following criteria: Library score > 0.75, RT shift < 1%, difference in isotope intensity < 10%, mass accuracy <5 ppm. **D** – Venn diagrams for the number of annotations in human urine and plasma in positive and negative ion mode (pH 3 and 8).

Conclusion

In-house processing tools allow ables single sample approach and fragments on product ion level and increases selectivity when possibility of adaption during data possibility of adaption during data acquisition processes
fragmentation pathway of co-eluting precursors

• Full workflow automation en-

 SWATH/MS allows reprocessing of all the data post-acquisition and reduces the rate of false analyte identification compared to SRM

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SWATH/MS acquisition enables quantification using any

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