

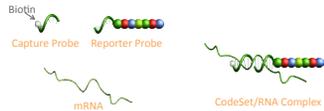
A Novel Multiplexed Digital Gene Expression Technology

Gary K. Geiss^{1,4}, Roger Bumgarner², Brian Birditt¹, Timothy Dahl¹, Naeem Dowidar¹, Dwayne L. Dunaway¹, H. Perry Fell¹, Sean Ferree¹, Renee D. George¹, Tammy Grogan¹, Jeffrey J. James¹, Malini Maysuria¹, Jeffrey D. Mitton¹, Paola Oliveri⁴, Jennifer L. Osborn³, Tao Peng², Amber L. Ratcliffe¹, Philippa J. Webster¹, Eric H. Davidson⁴, and Leroy Hood⁵

¹NanoString Technologies Inc., 201 Elliott Ave West, Suite 300, Seattle, WA 98119 ²The Department of Microbiology, Box 358070, University of Washington, Seattle WA 98195 ³Current address, Department of Bioengineering, Box 355061, University of Washington, Seattle WA 98195 ⁴Division of Biology 156-29, California Institute of Technology, Pasadena CA 91125 ⁵The Institute of Systems Biology, 1441 N. 34th St., Seattle WA 98103

TECHNOLOGY

One code = One mRNA

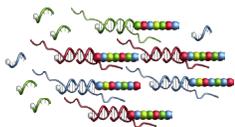


CodeSets are color coded "barcodes" that employ two 50-bp probes per mRNA which hybridize in solution. The Reporter Probe carries the signal, the Capture Probe allows the complex to be immobilized in the sample cartridge for data collection. Detection is direct and digital and the assay does not require cDNA synthesis or amplification.

WORKFLOW

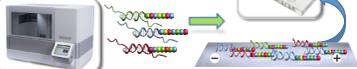
1 Hybridize

1. Buffer
2. CodeSet
3. Sample



Sample material is mixed with excess CodeSet and hybridized overnight.

2 Purify



The Prep Station removes excess CodeSet and immobilizes CodeSet/RNA complexes in the nCounter Cartridge for data collection.

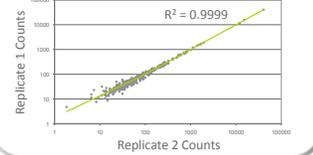
3 Count



Sample cartridges are placed in the Digital Analyzer for data collection. Images of color codes on the surface of the cartridge are counted, and a running total of each target is tabulated.

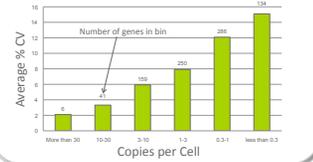
PERFORMANCE

Assay Sensitivity, Reproducibility and Dynamic Range



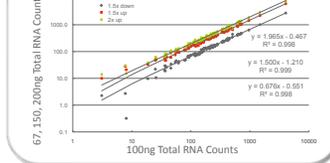
Transcript counts for two technical replicates of a 509-plex CodeSet demonstrate the assay reproducibility over a wide dynamic range. 75 counts equals a [c] of ~1 copy/cell of a transcript, illustrating the high level of sensitivity and precision of the assay even at very low levels of expression.

Precision Across Large Ranges



Targets for the 509-plex assay were grouped by level of expression to evaluate the precision of the assay across the dynamic range. Percent CV was calculated for each group. The number of targets for each group are indicated by the number above each column. Targets expressed at below a single copy per cell demonstrated CVs less than 15%.

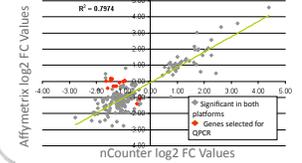
Fractional Fold Change Accuracy



The amount of input material was varied. The 100ng reaction was used as the baseline and 67, 150, and 200ng of input material was plotted against it. The slopes correlate closely with the expected values of 0.67, 1.5 and 2 which indicate the assay's ability to detect fractional fold changes.

CORRELATION

nCounter Analysis System vs. Affymetrix Microarray



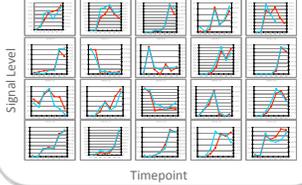
100ng of total RNA from Mock- and Polio Virus-infected cells were assayed for log₂ fold change using both nCounter and Affymetrix's U133 Plus 2.0 arrays.

Correlation between nCounter, Affymetrix & TaqMan



There was disagreement between nCounter and Affymetrix on 14 genes. TaqMan RT-PCR validated the nCounter results.

20 Genes Analyzed by SYBR Green and nCounter



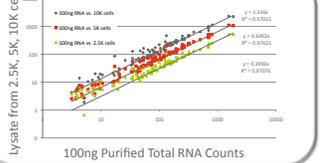
Expression levels for 20 sea urchin genes measured on the nCounter System at 7 development time points showed excellent correlation with SYBR Green Real Time PCR.

EFFICIENCY

Reaction Number Comparison Calculator	
Number of Samples	200
Number of Genes	200
Technical Replicates	3
Required #s of Reactions	nCounter: 600, qPCR: 120,000

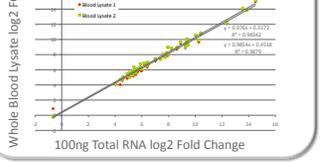
SAMPLE TYPES

Matched Cell Lysate and Total RNA Data



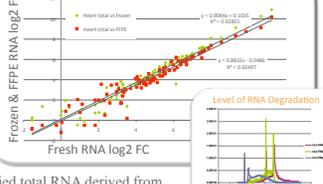
Raw cell lysate in varying amounts was input directly into the nCounter hybridization reaction and is compared here to a matched purified total RNA sample.

Matched Whole Blood Lysate and Total RNA Data



PaxGene lysed blood was input directly into the nCounter hybridization reaction and is compared here to a matched purified total RNA sample.

Matched Frozen and FFPE Derived Total RNA



Purified total RNA derived from matched fresh, frozen and FFPE treated tissue were assayed with the nCounter System.

SUMMARY

The NanoString assay offers many advantages including:

- Direct digital detection
- Multiplex hundreds of genes
- High sensitivity & precision
- No enzymology
- Fully automated
- Cost competitive

ACKNOWLEDGMENTS
We would like to acknowledge Edward Kelly, at the Center for DNA Sequencing and Gene Analysis, UW School of Pharmacy, Department of Pharmaceutics for TaqMan analysis and the University of Washington's Center for Array Technologies for processing of Affymetrix arrays.

www.nanostring.com

nanoString
TECHNOLOGIES