

Application of Central Composite Design (CCD) for the Optimization of Ultrasound-assisted Extraction (UAE) of Phenolics from Corn (Zea mays)



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INTRODUCTON

Corn (Zea mays) is the third most important crop worldwide and might serves as a staple food, especially in the developing countries.¹ It becomes more important attributable to naturally presented phenolic compounds.^{2,3}

Many studies showed that phenolics have some benefits including play a key role in anti-oxidative defences mechanisms,⁴ exhibit health-promoting effects,⁵ and inhibitory effects on mutagenesis and carcinogenesis.⁶

The development phenolics extraction may complicate due to its structural diversity as having multiple hydroxyl groups.⁷ Furthermore, their potent antioxidant activity leads to fast reaction with other components in the matrix.⁸ Hence, a new extraction method that effectively liberate phenolics from matrices and rapidly recover these compounds is required.

Ultrasound-assisted extraction (UAE) appears to be the most promising process to extract phenolics from plant samples due to the cavitation effect, thus aids the extraction solvent to penetrate rapidly into the plant cells and prevents degradation of phenolic compounds.

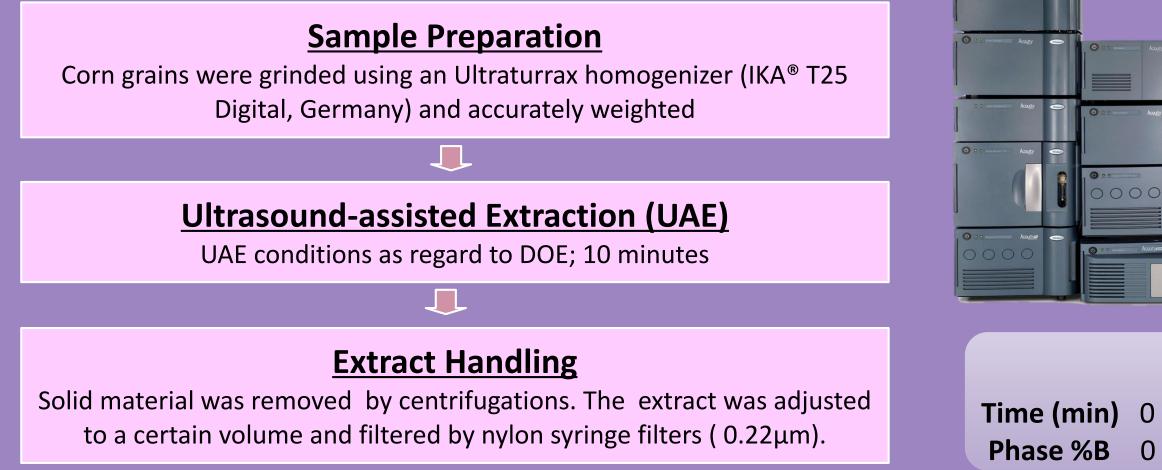
EXPERIMENTAL DESIGN

A central composite design (CCD) has been used to investigate the effects of six independent variables on the extraction yield at three levels. The range of independent factors and their levels is described bellow:

Variable	-1	0	+1	Unit	
Solvent (A)	0	25	50	%MeOH	
Temperature (B)	10	40	70	°C	urzoos and
Amplitude (C)	30	50	70	%	nietoter
Cycle (D)	0.2	0.45	0.7	-	
	2	5	7		

ANALYTICAL METHODS

The ultrasound-assisted extractions (UAE) were performed using an ultrasonic probe (model UP200S, medium probe, Hielscher Ultrasonics, 200 Watt).



The determination of phenolics was carried out on an ACQUITY UPLC[®] H-Class system coupled with a Photo-diode Array (PDA) detector and controlled by an Empower[™] 3 Chromatography Data Software (Waters Corporation, Milford, MA)

	Determination System									
	Flow Rate	:0.7 mL min ⁻								
	Injection Volume	: Ι0 μL								
	PDA scan range	: 200-400 nm								
	Column Temperature	:47 °C								
	Phase A : 0.01% Acetic A Phase B : 2% Acetic Acid									
Gradient Program										
Time (min) 0 1	1.1 2 3 3.5	4 7 8 10								

20

Estimated Response Surface

20

60 100 100 0

0

10

10

р**н** (Е) 3 3 / Ratio sample:solvent (F) 1:10 1:15 1:20

CCD consisted of 45 experimental points carried out in random order.



STATGRAPHICS[®] Centurion XVI (Statpoint Technologies, Inc., USA) has been used to evaluate the effects of operating parameters on the extracted yield. A secondorder model is applied in Response Surface Methodology (RSM):

 $y = \ \beta_0 + \ \sum_{i=1}^k \beta_i x_i + \ \sum_{i=1}^k \beta_{ii} x_i^2 + \ \sum_{i=1}^{k-1} \sum_{i=2}^k \beta_{ijx_ix_j} + \ \epsilon$

where $x_1, x_2, ..., x_k$ are the factors that influence the response y; β_0 , β_{ii} (i = 1, 2,..., k), β_{ij} (i = 1, 2, ...,k; j = 1, 2,...,k) are unknown parameters and ε is a random error. The β coefficients are obtained by the least square method.

RESURS

Analytical properties for the UPLC-PDA (Linear Range 0.15-12 mg L⁻¹)

		D 2	LOD	LOQ	Intra-day,	CV (%) n=9	Inter-day,	CV (%) n=3x3
Compounds	Linear Equation	R ²	(mg L ⁻¹)	(mg L ⁻¹)	RT	Area	RT	Area
Protocatechuic acid	y = 54421.70x + 208.24	0.9997	0.2	0.7	0.21	0.49	0.86	0.79
p-OHBenzoic acid	y = 116136.01x + 5629.27	0.9999	0.2	0.5	0.14	0.17	0.76	1.00
Vanillic acid	y = 95900.55x + 2976.36	0.9999	0.2	0.7	0.19	0.24	0.64	0.73
Chlorogenic acid	y = 13703.67x - 68448.81	0.9999	0.1	0.4	0.17	0.27	0.80	I.23
Caffeic acid	y = 108664.31x - 34804.94	0.9999	0.1	0.4	0.25	0.35	1.39	0.62
p-Coumaric acid	y = 166513.11x - 2772.38	0.9999	0.1	0.4	0.36	0.36	I.40	0.64
Ferulic acid	y = 141074.71x - 80282.13	0.9993	0.4	1.2	0.39	0.20	1.10	0.81

Additional optimization: Amplitude

a

b

1.5

1.3

1.1

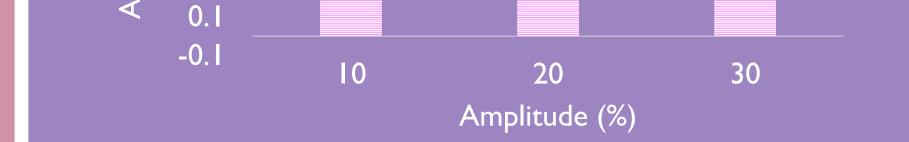
0.9

0.7

0.5

0.3

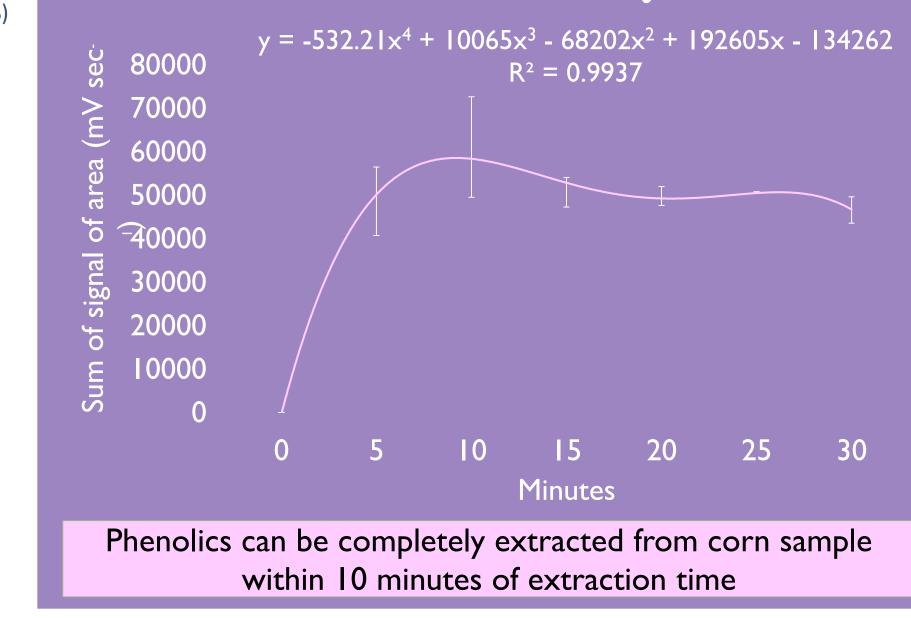
(280 nm)



(a)

Single-factor ANOVA indicates that amplitude has a significant effect as $F_{calculated}$ (170.6) >> $F_{critical}$ (5.14) on the extraction yield. Hence, 30% was defined as the optimum amplitude.

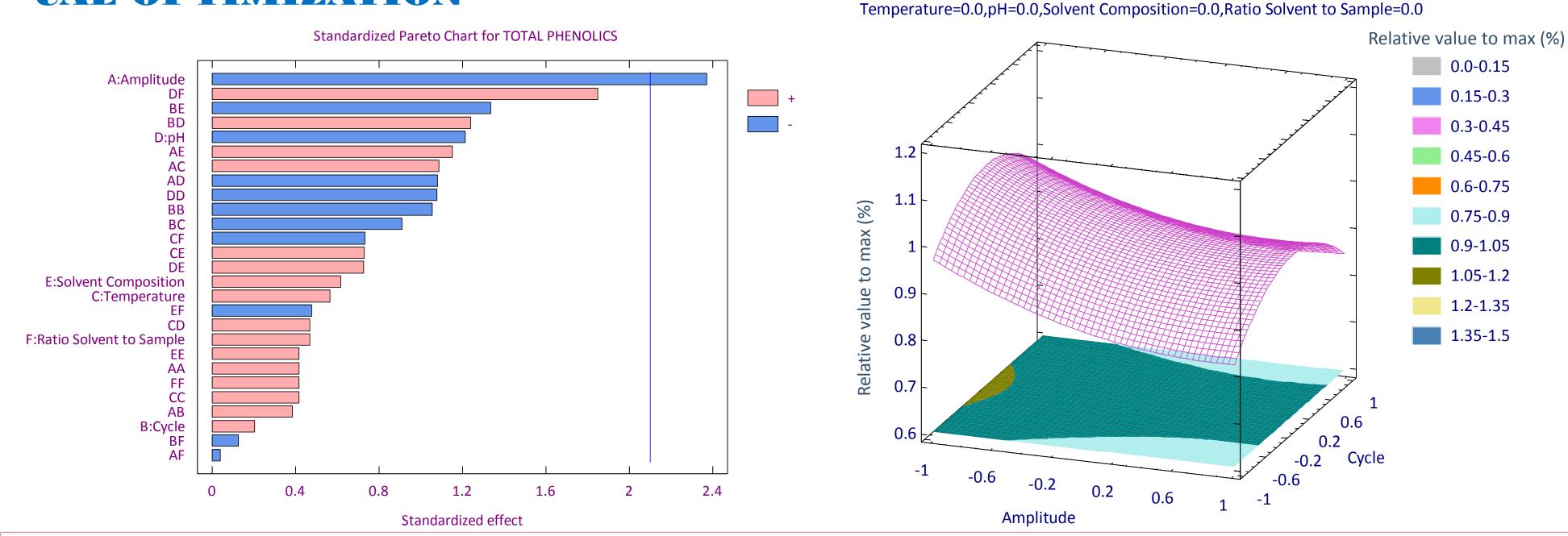
Kinetic Study



REAL SAMPLE APPLICATION

Red		Black	Yellow
	White	DIACK	Tentow
TR	ND	0.684±0.004	ND
ND	ND	ND	ND
0.701±0.023	ND	ND	TR
1.501±0.065	TR	0.588±0.196	TR
1.137±0.125	0.511±0.044	TR	0.623±0.011
1.190±0.047	0.784±0.053	1.558±0.002	0.446±0.049
1.619 <u>±</u> 0.192	1.534±0.060	TR	TR
	ND 0.701±0.023 1.501±0.065 1.137±0.125 1.190±0.047	ND ND 0.701±0.023 ND 1.501±0.065 TR 1.137±0.125 0.511±0.044 1.190±0.047 0.784±0.053	ND ND ND 0.701±0.023 ND ND 1.501±0.065 TR 0.588±0.196 1.137±0.125 0.511±0.044 TR 1.190±0.047 0.784±0.053 1.558±0.002

UAE OPTIMIZATION



Amplitude (A) significantly affects the recovery; \uparrow Amplitude $\equiv \uparrow \uparrow \uparrow$ Recovery Since the coordinates of the highest point of response was in the corner of the design region, further amplitude optimization should be tested.

Optimized conditions by

0.98

1.00

-1.00

-0.34

-0.17

-0.96

MeOH

Temp.

Cycle

pН

Ratio

Amplitude

PERFORMANCE OF THE METHOD

RSM Factors Optim Level

		Recovery (%)	Precisio		
num	m Compounds		Intra-day	Inter-day	
75%	Caffeic acid	71.51	1.02	2.13	
50°C	Ferulic acid	110.70	2.63	3.18	
30%	p-Coumaric acid	97.22	2.73	2.12),
0.4	<i>p</i> -OHBenzoic acid	73.50	1.05	1.14	
	Protocatechuic acid	84.33	3.80	10.57	
4.0	Sinapic acid	78.94	I.48	2.98	
I:2.5	Vanillic acid	72.19	9.81	12.01	



ne developed and validated ethod was then successfully applied to the analysis of enolics content in four corn Itivars including red, yellow, black and

Note: Unit in mg Kg⁻¹ sample ND, not detected due to less than LOD. TR, trace due to the concentration less than LOQ but higher than LOD.

The major phenolic compounds in corn grain were ferulic and **p-coumaric acid**. The grain from red corn cultivar consists the highest amount of phenolics while the yellow cultivar posses the lowest level of phenolic compounds.

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