Probabilistic Predictive Model of the Human Liver Microsomal Metabolism Regioselectivity

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As it could have been expected prior to

produce large numbers of false positive

this test, the baseline HLM models

enzymes. This is due to the fact that

metabolites produced by all enzymes,

each case can be only responsible for

some of them. As a result, the sensitivity

of HLM baseline models applied to the

individual enzyme data is comparable to

that of the HLM baseline model applied

to the HLM test set, while the specificity

is generally much lower (see Figure 2A).

corrections based on the Self-training

individual enzyme data, the number of

Libraries, containing corresponding

After the application of similarity

false positive atoms decreases

considerably. Within the Model

of metabolism in human liver

microsomes (see Figure 2B).

for an easy and straightforward

any drug metabolizing enzyme.

Applicability Domain (RI>0.3) the

specificity parameters for individual

enzyme regioselectivity predictions

become comparable to those of the

corresponding HLM data based models

Such results clearly illustrate that the

GALAS modeling methodology allows

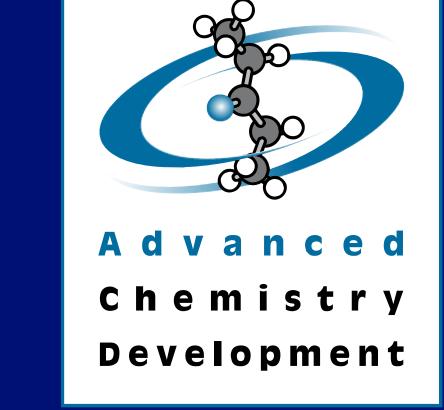
adaptation of the resulting HLM baseline

models for predicting regioselectivity of

yet a particular enzyme of interest in

predictions on data for separate

HLM models take into account



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INTRODUCTION

Analytical identification of metabolites for a drug candidate is usually a time consuming and low-throughput task which is performed only in late drug development phases. Therefore, the ability to predict possible sites of human liver microsomal metabolism using *in silico* techniques would be highly beneficial for any medicinal chemist. Moreover, *in silico* predictions of the most likely metabolism sites in a molecule could facilitate the analysis of spectroscopic data and thus ease the experimental identification of metabolites.

In this work, we present QSAR models for the prediction of metabolism regioselectivity. They provide the probability to be metabolized in human liver microsomes for every atom of the molecule and are based on a novel GALAS (Global, Adjusted Locally According to Similarity) methodology – an approach enabling the evaluation of Model Applicability Domain via the calculation of the prediction Reliability Index (RI).

DATA SET

Experimental data on metabolism in human liver microsomes for 873 compounds were collected from scientific publications dealing with analytical identification of the metabolites observed after the incubation of compound with human liver microsomes or recombinant cytochrome P450 enzymes. Every carbon atom with at least one hydrogen attached was marked as a site of metabolism, if

hydroxylation at the atom was observed, or

site of no metabolism otherwise. For dealkylation reactions, carbon atoms of the leaving groups were marked in the same manner. Similarly, every sulfur atom was marked whether oxidized or not. Some sites were marked as "inconclusive" and consequently not used in the modeling. The distribution of metabolized and not metabolized atoms for different enzymes is shown in Figure 1.

The complete dataset of ca. 8500 marked atoms was divided into aromatic hydroxylation, N-dealkylation, O-dealkylation, S-oxidation, and aliphatic hydroxylation subsets according to the atom type [2]. The composition of obtained subsets in case of human liver microsomal metabolism is outlined in Table 1. Each subset was further divided into training (70%) and test (30%) sets and individual models were built for all types of reactions.

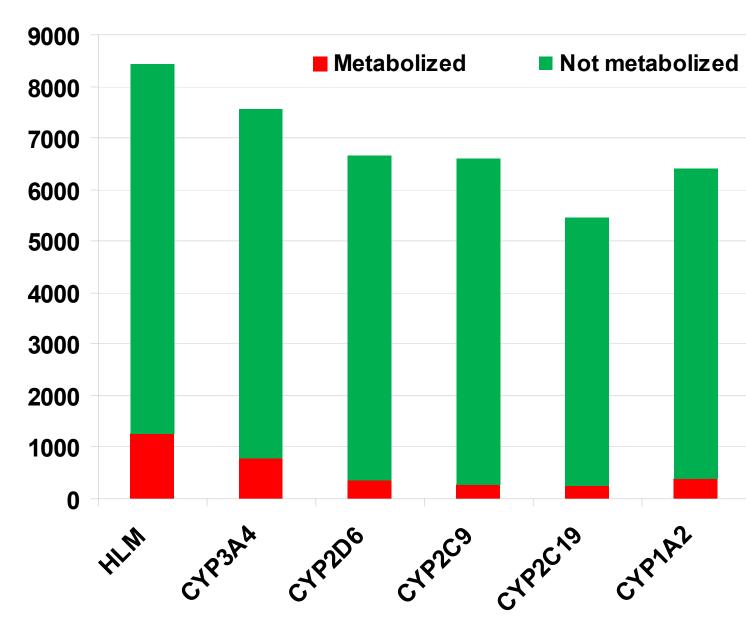


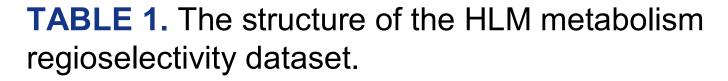
FIGURE 1. Distribution of metabolized and nonmetabolized atoms in datasets for cytochrome P450 enzymes and overall HLM metabolism.

MODEL DEVELOPMENT

The modeling workflow is shown in Scheme 1. The procedure consists of the following steps:

- Atom centered fragmentation for every marked atom;
 Development of a baseline model predicting the probability to be metabolized for every atom in a molecule using PLS in combination with bootstrapping;
- Correction of baseline predictions according to experimental data for 5 most similar atoms in the training set accompanied by the estimation of prediction reliability in the form of Reliability Index (RI)

The entire process of the molecule fragmentation and subsequent statistical analysis was realized using Algorithm Builder application [2]. More details on GALAS modeling method can be found in our recent publications [3,4].



Subset	No. of compounds	No. of metabolism sites	Total No. of marked atoms	
N-dealkylation	511	333	1173	
O-dealkylation	488	260	1033	
Aliphatic hydroxylation	723	318	2904	
Aromatic hydroxylation	739	358	3341	
S-oxidation	135	57	157	
Total	873	1326	8606	

Atom-centered fragmentation

Binomial PLS + Multiple Bootstrapping

 $X = Baseline logit(p) + \Delta logit(p)$

SCHEME 1: Outline of modeling

methodology

Multiple predictors

Coefficients (a_i)

 $logit(p) = \sum a_i x$

Omeprazole (C8)
Omeprazole (C20

olbutamide (C15)

Library

ctions, carbon atoms of the leaving groups were tom was marked whether oxidized or not. Some sites used in the modeling. The distribution of metabolized hown in Figure 1.

*- Unreliable predictions (RI<0.3) were not considered which led to exclusion of 73 marked atoms (21 metabolism sites) test set in case of N-dealkylation. 66 marked atoms (26 metabolism sites) in case of O-dealkylation. 111 marked atoms (27 metabolism sites).

The baseline models already provide satisfactory prediction results, however some problems are also evident. For example, in case of aliphatic and aromatic hydroxylations, a large number of false positive predictions is observed (~20% of all predictions). This situation noticeably improves after the application of similarity correction. Analogous improvement of baseline model predictions is also obtained in case of N- and O-dealkylation models. The percent of correct classifications produced by these models approaches 100 if only predictions of high reliability (RI>0.5), which still constitute ca. 50% of the test set, are taken into account. These results prove the effectiveness of local similarity correction and the usefulness of Reliability Indices in assessing the Model Applicability Domain as the statistical parameters of the model clearly correlate with the reliability of considered predictions.

MODEL ADAPTATION FOR INDIVIDUAL CYTOCHROME P450 ENZYMES REGIOSELECTIVITY PREDICTION

Table 3 below presents results for a few selected examples of a GALAS model training with separate enzyme data. The baseline model for the prediction of each particular reaction regioselectivity in HLM was separately trained with a corresponding reaction data for each of the five considered individual P450 enzymes (e.g., in Table 3 HLM N-dealkylation baseline model was trained using CYP1A2 N-dealkylation data, HLM baseline model for the prediction of aromatic hydroxylation – using CYP2D6 aromatic hydroxylation data, etc.).

TABLE 3. Results for a few selected examples of an HLM model adaptation using separate enzyme data.

			Predicted value						
			Baseline*		Similarity Corrected*		Similarity Corrected (RI>0.5)		
			Positive	Negative	Positive	Negative	Positive	Negative	
	CYP1A2 N-dealkylation	Positive	14	5	13	6	4	3	
		Negative	25	145	7	162	3	73	
ne	CYP2C9 O-dealkylation	Positive	6	1	5	2	2	0	
Observed value		Negative	30	135	8	157	1	77	
	CYP3A4 Aliphatic hydroxylation	Positive	31	17	28	20	18	8	
		Negative	136	538	23	651	5	453	
	CYP2D6 Aromatic hydroxylation	Positive	21	1	17	5	11	1	
		Negative	176	512	15	673	5	446	

^{* -} Unreliable predictions (RI<0.3) were not considered which led to exclusion of 74 marked atoms (8 metabolism sites) from the initial test set in case of CYP1A2 N-dealkylation, 56 marked atoms (11 metabolism sites) in case of CYP2C9 O-dealkylation, 84 marked atoms (16 metabolism sites) in case of CYP3A4 aliphatic hydroxylation and 65 marked atoms (10 metabolism sites) in case of the CYP2D6 aromatic hydroxylation.

MODEL VALIDATION

TABLE 2. Detailed results of the HLM metabolism regioselectivity algorithms' validation using the test sets constituting 30% of the initial HLM databases for each reaction.

			Predicted value						
			Baseline*		Similarity Corrected*		Similarity Corrected (RI>0.5)		
			Positive	Negative	Positive	Negative	Positive	Negative	
	N-dealkylation	Positive	48	31	60	19	33	5	
		Negative	10	190	10	190	3	105	
	O-dealkylation	Positive	44	11	46	9	23	1	
ne		Negative	10	179	7	182	3	118	
ed value	Aliphatic hydroxylation	Positive	41	21	35	27	27	11	
Observed		Negative	149	574	31	692	13	498	
	Aromatic hydroxylation	Positive	72	25	56	41	33	17	
		Negative	191	669	46	814	22	589	
	S-oxidation	Positive	7	0	6	1	3	0	
		Negative	1	20	1	20	0	16	

* - Unreliable predictions (RI<0.3) were not considered which led to exclusion of 73 marked atoms (21 metabolism sites) from the initial test set in case of N-dealkylation, 66 marked atoms (26 metabolism sites) in case of O-dealkylation, 111 marked atoms (27 metabolism sites) in case of aliphatic hydroxylation, and 84 marked atoms (14 metabolism sites) in case of the aromatic hydroxylation, and 20 marked atoms (13 metabolism sites) in case of S-oxidation.

PREDICTION EXAMPLES

N-dealkylation O-dealkylation Aliphatic hydroxylation

HLM baseline model applied on test sets of corresponding individual enzymes (Table 3 – Baseline column)

B

N-dealkylation O-dealkylation Aliphatic hydroxylation

Aliphatic hydroxylation individual enzymes (Table 3 – Baseline column)

N-dealkylation O-dealkylation Aliphatic hydroxylation hydroxylation

HLM baseline model with HLM training set as a Self-training Library applied on HLM test set (Table 2 – Similarity corrected column)

HLM baseline model with individual enzyme datasets as Self-training Library applied on test sets of corresponding individual enzymes (Table 3 – Similarity corrected column)

FIGURE 2. Specificity of metabolism regioselectivity models.

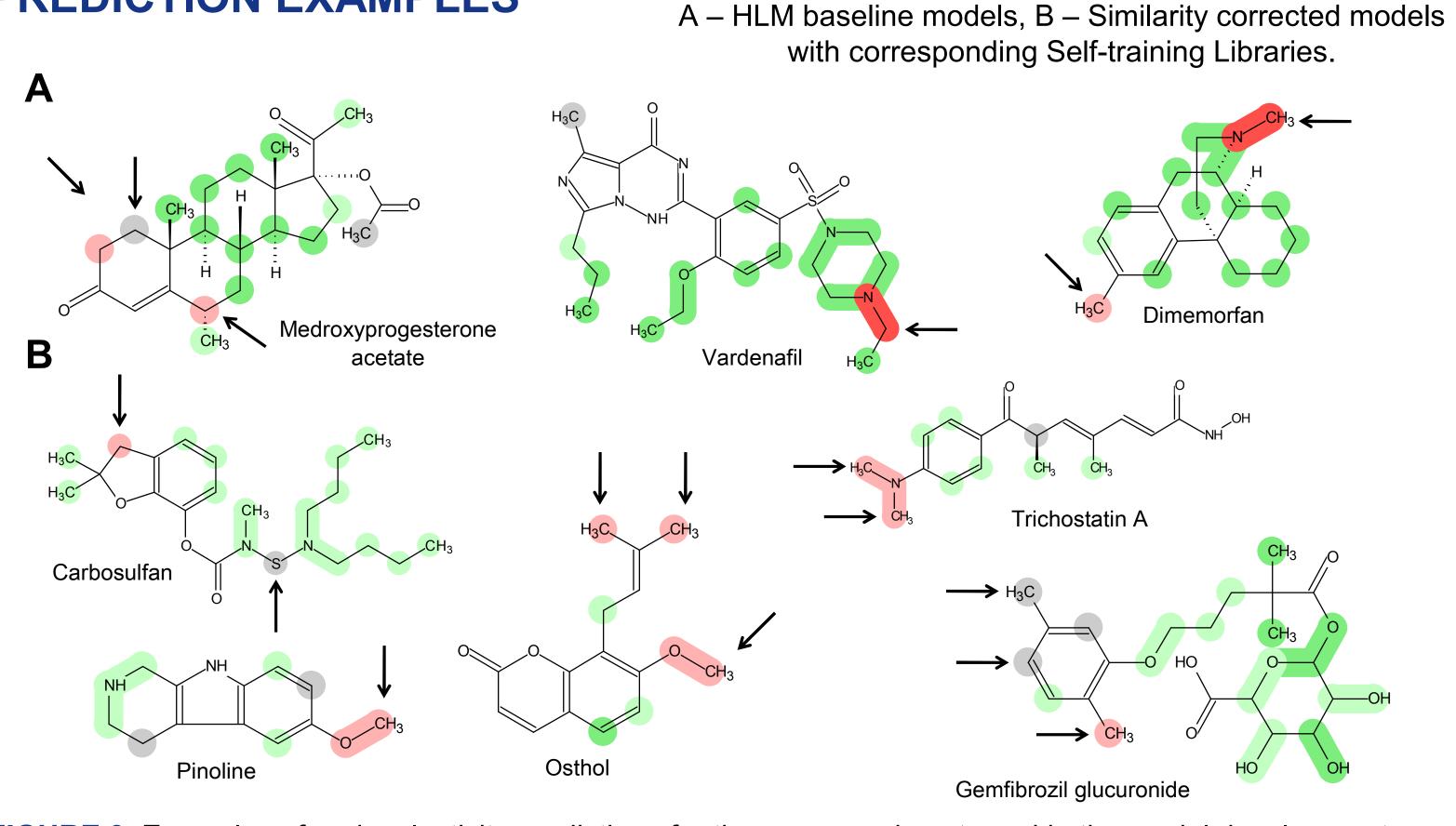


FIGURE 3. Examples of regioselectivity predictions for the compounds not used in the model development: A – compounds from well-known drug classes, B – novel drug-like molecules having nothing similar in the training set of the model. Arrows indicate experimentally identified metabolism sites.

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