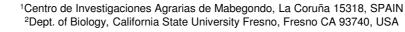


DNA polymorphism in the pepper-Phytophthora capsici pathosystem

Cristina Fernández-Otero^{1,2}, Jesús Moreno-González¹, and James P. Prince²





Introduction

Phytophthora capsici is a pathogen on several economically important crops, affecting tomatoes, eggplants, squash, and melons, and it is one of the most damaging pathogens currently affecting pepper (*Capsicum annuum*) worldwide (Tyler 2002, Oelke *et al.* 2003). The species *Phytophthora capsici* was first reported in 1922 in New Mexico and spread to vegetable production areas in Colorado and Florida in the 1930's and 1940's. This oomycete attacks the roots, stems, leaves and fruit of the plant. Resistance in *Capsicum annuum* is genetically complex. In order to integrate genetic linkage maps of peppers and P. capsici, a series of SSRs, RAPD, CAPS and COSII markers and candidate genes for virulence (glucanase, pectinase, chitinase and cutinase) have been tested in a panel of 6 different C. annuum (American and European genotypes) and in 2 P. capsici selected because they are the parents of a segregating cross used for linkage mapping. We are interested in finding polymorphisms in the pepper-P. capsici system because these could serve as potential genetic markers for the construction of linkage maps.

Materials and Methods

Population

Six pepper (C. annuum) genotypes were used: "Padrón 124", "Couto 12B", "CM334", "PI201234", "PSP-11" and "NuMex Joe E. Parker (JEP)", with different levels of resistance to P. *Capsici*. "CM334" is the most resistant according to previous research (Guerrero and Laborde 1980, Pochard et al. 1983, Gil Ortega et al. 1992)

Two P. capsici isolates, Ppc3 and GSP1-1, selected for different levels of virulence on pepper, were used.

<u>DNA extractions</u> DNA genomic was extracted following the procedures of Prince *et al.* 1997, and using the "Mini-Plant Genomic DNA Isolation Kit " (Metabion)

Molecular markers

RAPD: were generated using 20 different decamers (Operon Technology, Alameda, Calif.)

CAPS: primer pairs have been obtained based on previously sequenced and mapped tomato RFLP clones, chosen because of their even distribution throughout the genome of tomato. COSII: chosen because of their distribution in tomato and Arabidopsis. Sequences available at

ftp://ftp.sgn.cornell.edu/COSII/pepper_mapping/ SSRs: P. capsici EST sequences from the Phytophthora Functional Genomics Database (Gajendran et al.

2006) were screened for SSR sequences using Sputnik (Abajian 1994). Candidate genes for virulence: Primers sets have been designed from previously cloned genes that we are investigating as potential candidate genes for virulence: glucanase, pectinase, chitinase and cutinase. We used various of the above molecular markers to detect polymorphism on C. annuum and P. capsici.

PCR amplicons were analyzed on 1- 1.5% (w/v) agarose gel.

Results

More than 75 molecular markers have been designed and used to analyze polymorphism in the

P. capsici- pepper pathosystem, so far. Fifteen polymorphisms were found in pepper plants and nineteen between the two isolates of *P. capsici* studied (see Tables 1 and 2).

We detected the highest levels of polymorphism (75%) in *P. capsici* using candidate genes for virulence and in pepper plants using RAPD markers (>65%). Polymorphisms in the peppers seem to be slightly more common among JEP and CM334 than between the rest

of the pairs of mapping parent genotypes (data not shown). (Figures 1 and 2 show examples of polymorphism found using the different types of molecular markers).

	Phytophthora capsici	
Type of Marker	Number Available for screening	Number Polymorphic
SSR	13	2
RAPD	20	10
Virulence genes	4	3
TOTAL	37	15

	Capsicum annuum	
Type of Marker	Number Available for screening	Number Polymorphic
COSII	32	10
RAPD	9	6
CAPS	6	3
TOTAL	47	19

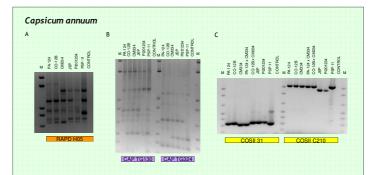
Table 1 and 2. Molecular markers being used in P. capsici and Capsicum annuum screening

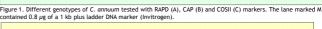


Acknowledgements

We would like to thank to Dylan Storey and Mitch Lucas for their help in the RAPD, CAP and COSII markers selection.

C. F-O is a postdoctoral researcher from Xunta de Galicia and acknowledges the receipt of a fellowship during the undertaking of the present study in US. The CSU Fresno College of Science & Mathematics has partially supported this work





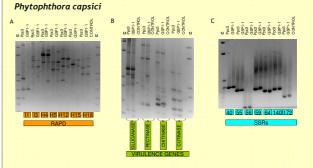


Figure 2. Different genotypes P capsici tested with RAPD markers (A), candidate genes for virulence (B) and COSII markers (C). The lane marked M contained 0.8 μ g of a 1 kb plus ladder DNA marker (Invitrogen).

References

Abajian, C. 1994. Sputnik. http://espressosoftware.com/pages/sputnik.jsp

Gajendran K, Gonzales MD, Farmer A, Archuleta E, Win J, Waugh ME, and Kamoun S. 2006. Phytophthora functional genomics database (PFGD): functional genomics of Phytophthora-plant interactions. Nucl. Acids Res. 34:D465-D470.

Gil Ortega R, Palazón Español C, Cuartero Zueco J. 1992. Genetic relationships among four pepper genotypes resistant to *Phytophthora capsici*. Plant Breeding 108: 118-125.

Guerrero-Moreno A and Laborde JA. 1980. Current status of pepper breeding for resistance to Phytophthora capsici in Mexico. In: Proc IVth Eucarpia Meeting on Capsicum. Wageningen, The Netherlands. pp 52-56.

Oelke LM, Bosland PW, and Steiner R. 2003. Differentiation of race specific resistance to Phytophthora root rot and foliar blight in Capsicum annuum. J. Am. Soc. Hortic. Sci. 128:213-218.

Pochard E, Molot P and Dominguez G. 1983. Étude de deux nouvelles sources de résistance à Phytophthora capsici Leon. chez le piment: confirmation de l'existence de tríos composantes distinctes dans la résistance. Agronomie 3: 333-342.

Prince JP, Zhang Y, Radwanski ER and Kyle MM. 1997. A versatile and high-yielding protocol for the preparation of genomic DNA from *Capsicum* spp. (pepper). Hort Sci. 32: 937-939. Tyler B M. 2002. Molecular basis of recognition between Phytophthora pathogens and their hosts. Ann.

Rev. Phytopath, 40:137-167